

BBA 45913

Mn²⁺ AS ELECTRON DONOR IN ISOLATED CHLOROPLASTS

G. BEN-HAYYIM AND M. AVRON

Biochemistry Department, Weizmann Institute of Science, Rehovot (Israel)

(Received October 16th, 1969)

SUMMARY

1. Added Mn²⁺ is concluded to serve as an electron donor at a site which precedes Photosystem 2. This conclusion is supported by the following observations: (a) Addition of Mn²⁺ increased the apparent rate of O₂ uptake in the presence of 1,1'-ethylene-2,2'-dipyridilium dibromide. (b) Mn²⁺ had little or no effect on NADP⁺ photoreduction, while the concomitant O₂ evolution was severely inhibited. (c) All the reactions in which Mn²⁺ served as the electron donor were as sensitive to 3-(3,4-dichlorophenyl)-1,1-dimethylurea as the Hill reaction and were insensitive to CN⁻ or N₃⁻.

2. In the presence of Mn²⁺ and Fe(CN)₆³⁻ high ratios of P/2e were obtained indicating the existence of a cyclic process due to an interaction between the products of the reaction, Fe(CN)₆⁴⁻ and Mn³⁺.

3. A method was developed for assaying the Mn³⁺ formed. In the presence of both Mn²⁺ and water as electron donors to Photosystem 2, the Mn³⁺ produced accounted for about 80 % of the electron flow observed.

INTRODUCTION

The role of manganese in photosynthetic organisms is one of the longest studied aspects of photosynthesis. Studies concerning photosynthesis and Hill reaction in manganese-deficient whole plants, algae or chloroplast preparations revealed an absolute requirement for this nutrient¹⁻¹⁰. With the concept of two light reactions it was possible to demonstrate that the manganese is essential for Photosystem 2-mediated reactions. Thus, O₂ evolution^{11,12} or fluorescence properties¹³⁻¹⁶ which are dependent on Photosystem 2 activity were inhibited in the absence of manganese and the activity was restored by its readdition^{17,18}. On the other hand, KESSLER^{5,19} showed that photoreduction of CO₂ by hydrogen in hydrogen-adapted algae was not affected by manganese deficiency. This reaction was shown to require only Photosystem 1 activity. Thus, manganese is not an essential cofactor in reactions mediated by this photosystem. ANDERSON *et al.*²⁰ found that the fractionation of chloroplasts by treatment with digitonin yielded two fractions, a light and a heavy fraction corresponding to Photosystem 1 and Photosystem 2, and manganese was found to be tightly bound to the heavy particle.

Abbreviations: diquat, 1,1'-ethylene-2,2'-dipyridilium dibromide; DCMU, 3-(3,4-dichlorophenyl)-1,1-dimethylurea; HQNO, 2-*n*-heptyl-4-hydroxyquinoline-*N*-oxide.

However, despite ample evidence for a manganese requirement in photosynthetic reactions no report of light-induced changes in endogenous manganese have appeared. It is generally postulated that manganese serves as a catalyst in one of the electron transport steps of the O₂-evolving complex.

Several authors observed that exogenous Mn²⁺ was photooxidized by isolated chloroplasts. KENTEN AND MANN^{21,22} were the first to show photooxidation of Mn²⁺ to Mn³⁺ and they were followed by others^{23,24}. In all these studies it was suggested that Mn²⁺ could serve as an electron donor for Photosystem 2. However, all could not rule out the possibility of an indirect oxidation of Mn²⁺, namely by the H₂O₂ which was produced under the reaction conditions employed (see DISCUSSION in ref. 23).

In the present study we examined the effect of added Mn²⁺ on various types of Hill reactions. By introducing NADP⁺ or Fe(CN)₆³⁻ as electron acceptors rather than viologens or FMN, we could exclude any possible reaction between Mn²⁺ and H₂O₂. We investigated the effect of Mn²⁺ on electron transport to 1,1'-ethylene-2,2'-dipyridilium dibromide (diquat), NADP⁺ and Fe(CN)₆³⁻, its site of action and the rate of its photooxidation. Mn²⁺ was found to interact directly with the electron transport chain serving as electron donor to Photosystem 2.

METHODS

Chloroplasts were isolated from lettuce leaves as previously described²⁵. Chlorophyll was assayed after ARNON²⁶. O₂ uptake and evolution were measured with a Yellow Springs Instruments Clark type oxygen electrode. NADP⁺ photoreduction was measured in a Cary 14 recording spectrophotometer equipped with a scattering attachment. The photomultiplier was protected from the actinic light by 1 cm of a saturated solution of CuSO₄ and a corning C.S. 7-60 filter. Illumination was provided by a 500-W projector and was filtered through Baird Atomic sharp cut-off interference filters (all blocked to infinity) peaking at 640 mμ (25-mμ half-band width) and 715 mμ (15-mμ half-band width). When monochromatic light was not required a corning C.S. 2-58 filter was used. The intensity of the actinic light was measured with a Yellow Springs Instrument Radiometer Model 65 and was varied by using calibrated metal screens. Absorption data were calculated from a curve previously published²⁷.

Fe(CN)₆³⁻ photoreduction was measured either by following the decrease in absorbance at 420 mμ in a Cary 14 recording spectrophotometer or with an Aminco-Chance dual-wavelength spectrophotometer following the change 420—470 mμ. The photomultiplier was protected from the actinic light by a corning C.S. 4-96 filter. ATP formation was assayed as previously described²⁵.

Oxidation of Mn²⁺ to Mn³⁺ was measured by reduction of the Mn³⁺ to Mn²⁺ with Fe(CN)₆⁴⁻ followed by a sensitive determination of the remaining Fe(CN)₆⁴⁻. MCKENNA AND BISHOP²⁴ utilized a similar principle for standardization of Mn₄(P₂O₇)₃ but found their assay inadequate for estimation of Mn³⁺ formed by illuminated chloroplasts. The procedure used in this paper was as follows: A reaction mixture containing Mn²⁺ and pyrophosphate (to stabilize the oxidized Mn³⁺ as Mn₄(P₂O₇)₃) was illuminated for the desired period. 3.0 ml of the suspension were added to 1.0 ml of 1 mM Fe(CN)₆⁴⁻ solution. After incubation of 3–5 min in the dark at room tem-

perature, 0.4 ml of 30% trichloroacetic acid was added to inactivate the system. The remaining $\text{Fe}(\text{CN})_6^{4-}$ in the clear supernatant was determined by the method of AVRON AND SHAVIT²⁸.

RESULTS

Addition of Mn^{2+} to illuminated chloroplast preparations which consume O_2 in the presence of diquat resulted in an increase of the apparent rate of the O_2 uptake (Fig. 1a). Similar effects were observed when the Mn^{2+} was added to an uncoupled system (Fig. 1b). In both cases, the O_2 uptake was as sensitive to 3-(3,4-dichlorophenyl)-1,1-dimethylurea (DCMU) as was the $\text{Fe}(\text{CN})_6^{3-}$ Hill reaction. The effect of Mn^{2+} as a function of its concentration is shown in Fig. 2. As will be discussed later, the increase of the apparent rate of O_2 uptake does not seem to be due to an increase in the rate of electron transport but rather to the fact that Mn^{2+} served as electron donor prior to Photosystem 2. The stoichiometry for calculating

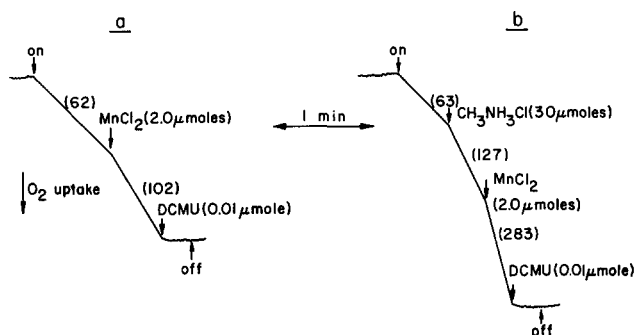


Fig. 1. Effect of Mn^{2+} on O_2 uptake in the presence of diquat. Reaction mixture contained: Tricine (pH 8.0), 45 μmoles ; NaCl, 60 μmoles ; diquat, 0.03 μmole ; NaN_3 , 3 μmoles and chloroplasts containing 73 μg chlorophyll, in a total volume of 3.0 ml. Numbers in parentheses represent rates of O_2 uptake expressed in μmoles per mg chlorophyll per h.

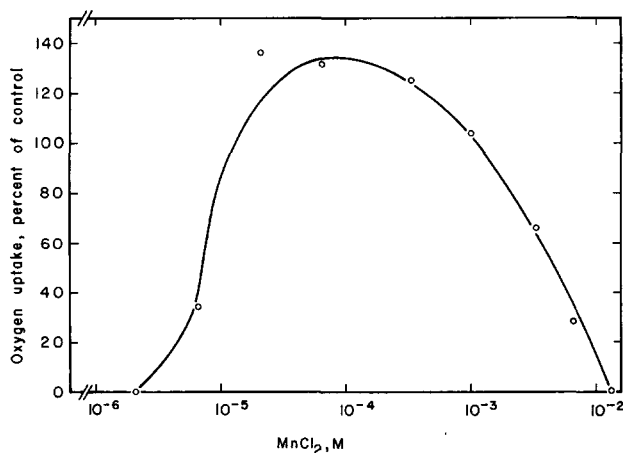
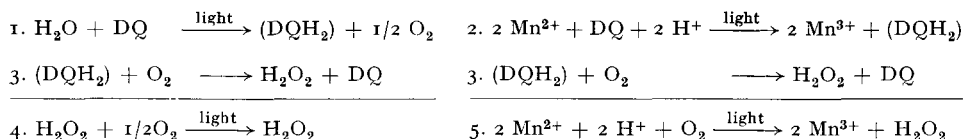


Fig. 2. Concentration curve for the effect of Mn^{2+} . Reaction mixture as described for Fig. 1. The control was measured in the absence of MnCl_2 , its activity being 18 μmoles O_2 per chlorophyll per h. Chloroplasts contained 100 μg chlorophyll.

the electron transport rate in the absence and presence of Mn²⁺ can be depicted by Eqns. 1, 3, 4 and 2, 3, 5, respectively.



Thus one molecule of O₂ taken up represents the photoinduced transfer of 4 electrons in the absence of Mn²⁺ but of only 2 electrons in the presence of Mn²⁺. With DQ as an electron acceptor O₂ is involved both at the donor and the acceptor sites. By using NADP⁺ as an electron acceptor one can measure separately the effect of Mn²⁺ on the donor and the acceptor ends of the electron transport chain. As seen in Fig. 3, the effect of Mn²⁺ on NADP⁺ photoreduction was different from that on O₂ evolution. While the O₂ evolution was strongly inhibited, NADP⁺ photoreduction stayed essentially the same.

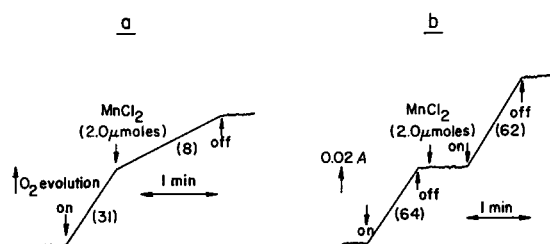


Fig. 3. Effect of Mn²⁺ on electron transport to NADP⁺. Reaction mixture contained: Tricine (pH 8.0), 45 μmoles; NaCl, 60 μmoles; NADP⁺, 0.5 μmole; saturating amount of ferredoxin and chloroplasts containing 65 μg chlorophyll in 3.0 ml. NADP⁺ photoreduction was followed by increase in absorbance at 350 mμ in a Cary 14 spectrophotometer as described under METHODS. Numbers in parentheses are initial rates of O₂ evolution and NADP⁺ photoreduction expressed in μmoles O₂ per mg chlorophyll per h and μmoles NADPH per mg chlorophyll per h, respectively.

The effect of Mn²⁺ on the electron transport to either diquat or NADP⁺ was highly specific for these ions. Mg²⁺ or Ca²⁺ at the same concentration was totally inactive or even slightly inhibitory. As seen in Table I, Mn²⁺ increased the apparent rate of O₂ uptake mediated by diquat to about the same extent as it decreased the O₂ evolution concomitant to NADP⁺ photoreduction. Under all tested conditions it was not possible to fully inhibit the O₂ evolution accompanying NADP⁺ reduction by the addition of Mn²⁺.

The site of action of Mn²⁺

The sensitivity of the Mn²⁺ effect to DCMU indicated that its site of action was relatively close to Photosystem 2. Quantum requirement measurements of NADP⁺ photoreduction in the presence and absence of Mn²⁺ exhibited the 'red drop' phenomenon characteristic of light-induced reactions in which Photosystem 2 participates.

As can be seen in Table II, Mn²⁺ somewhat lowered the quantum requirement at 640 mμ but did not change the quantum requirement at 715 mμ. Therefore, the

'red drop' in the presence of Mn^{2+} became even more pronounced than that with water.

Mn^{2+} failed to reactivate NADP^+ photoreduction in Tris-treated chloroplast preparations, possibly indicating that their site of action not only precedes Photosystem 2 but is rather close to the O_2 evolution step itself.

TABLE I

THE SPECIFICITY OF THE EFFECT OF Mn^{2+} ON PHOTOREACTIONS TAKING UP OR EVOLVING O_2

Reaction mixture contained: Tricine (pH 7.8), 45 μmoles ; NaCl, 60 μmoles ; chloroplasts containing 70 μg chlorophyll in a total volume of 3.0 ml. 0.03 μmole diquat and 3 μmoles NaN_3 were added in the reaction with diquat and 0.5 μmole NADP^+ with a saturating amount of ferredoxin were added in the NADP^+ reaction. The activity of O_2 evolution with NADP^+ and O_2 uptake in presence of diquat and in absence of Mn^{2+} (taken as 100%) were 20 and 44 $\mu\text{moles O}_2$ per mg chlorophyll per h, respectively.

Electron acceptor	Ion added	Activity (% of control)
NADP^+	None	(100)
	Mn^{2+}	36
	Mg^{2+}	100
Diquat	None	(100)
	Mn^{2+}	170
	Mg^{2+}	71
	Ca^{2+}	82

TABLE II

QUANTUM REQUIREMENT OF NADP^+ PHOTOREDUCTION IN THE PRESENCE OF Mn^{2+}

Reaction mixture contained: Tricine (pH 7.8), 45 μmoles ; NaCl, 60 μmoles ; aminomethane·HCl, 18 μmoles ; NADP^+ , 1 μmole ; saturating amount of ferredoxin and chloroplasts containing 39 μg chlorophyll in 3.0 ml. 2.0 μmoles of Mn^{2+} were added where indicated. NADP^+ photoreduction was measured as described under METHODS. Quantum requirement data represent values extrapolated to zero intensity (see ref. 27).

Electron donor	Quanta/electron		Ratio 715 m μ /640 m μ
	640 m μ	715 m μ	
H_2O	5.1	12.6	2.5
Mn^{2+}	3.7	14.2	3.8

The photoreduction of NADP^+ in the presence of Mn^{2+} was as sensitive as NADP^+ Hill reaction to all the inhibitors tested including DCMU, *o*-phenanthroline, ioxynil and 2-*n*-heptyl-4-hydroxyquinoline-*N*-oxide (HQNO).

In contrast to NADP^+ photoreduction, Mn^{2+} inhibited both $\text{Fe}(\text{CN})_6^{3-}$ photoreduction and the concomitant O_2 evolution to the same degree. The inhibition of O_2 evolution was believed to arise from the competition between Mn^{2+} and water, in a manner similar to that described for the case of NADP^+ . However, since (as will be shown later) Mn^{2+} is photooxidized to Mn^{3+} , this Mn^{3+} is rereduced by the $\text{Fe}(\text{CN})_6^{4-}$ which is formed when $\text{Fe}(\text{CN})_6^{3-}$ is the electron acceptor. Since it is impossible to separate the two processes, we studied the $\text{Fe}(\text{CN})_6^{3-}$ -dependent photo-

phosphorylation as an indirect measurement of the rate of electron flow from either water or Mn²⁺ to Fe(CN)₆³⁻. If indeed a cyclic electron flow of the type just described occurs, one would expect the rate of ATP formation to remain constant while the apparent rate of Fe(CN)₆³⁻ reduction is curtailed, resulting in an apparent increase in the ATP/2e ratio. As shown in Table III, this was indeed the case. As can be expected, the effect was most striking at the lower Fe(CN)₆³⁻ concentrations, where the Fe(CN)₆⁴⁻ to Fe(CN)₆³⁻ ratio would be highest.

Photooxidation of Mn²⁺

The Mn²⁺ effect on electron transport was interpreted as a result of Mn²⁺ serving as an electron donor. If so, oxidation of Mn²⁺ should occur. Such oxidation, of Mn²⁺ to Mn³⁺, was already demonstrated by others^{23,24} though at very low rates. Mn³⁺ in aqueous solution is extremely unstable, but addition of pyrophosphate to form Mn₄(P₂O₇)₃ yields a more stable complex. A method for assaying Mn³⁺ was developed (see METHODS) which enabled us to follow a time-course of the photooxidation of Mn²⁺ in a chloroplast preparation which consumed O₂ in the presence of diquat. NaN₃ was replaced by ethanol and catalase to avoid any possible complication due to a chemical reaction between Mn³⁺, peroxide and the added Fe(CN)₆⁴⁻. As

TABLE III

APPARENT P/2e RATIOS OF Fe(CN)₆³⁻ PHOTOREDUCTION IN THE PRESENCE OR ABSENCE OF Mn²⁺

Reaction mixture contained: Tricine (pH 8.0), 45 μmoles; NaCl, 60 μmoles; MgCl₂, 12 μmoles; phosphate buffer (pH 8.0), 12 μmoles (containing 9 · 10⁶ counts/min ³²P; ADP, 4 μmoles; chloroplasts containing 43 μg chlorophyll and 2.0 μmoles MnCl₂ where indicated, in a total volume of 3.0 ml. Fe(CN)₆³⁻ reduction was measured in a dual-wavelength spectrophotometer as described under METHODS. Each sample was illuminated for 2 min and later analyzed for its ATP content.

Initial Fe(CN) ₆ ³⁻ concn. (μM)	ATP formation (μmoles per mg chlorophyll per h)		Apparent P/2e	
	-Mn ²⁺	+Mn ²⁺	-Mn ²⁺	+Mn ²⁺
500	292	240	1.0	2.5
170	167	134	1.1	2.5
50	58	52	1.0	4.3
17	26	17	1.8	7.9

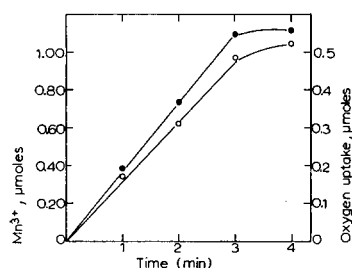


Fig. 4. Photooxidation of Mn²⁺ by isolated chloroplasts. Reaction mixture contained: Tricine (pH 7.8), 90 μmoles; NaCl, 120 μmoles; diquat, 0.1 μmole; Na₄P₂O₇ (pH 7.8), 60 μmoles; ethanol, 1020 μmoles; catalase, 100 μg; MnCl₂, 4 μmoles and chloroplasts containing 129 μg chlorophyll in 6.0 ml. Mn³⁺ content was determined as described under METHODS. The O₂ uptake in absence of Mn²⁺ was 0.14 μmole for the first 2 min. Full dots indicate O₂ uptake.

can be seen in Fig. 4, the photooxidation of Mn^{2+} was linear with time during the first few minutes and it accounted for about 80 % of the O_2 uptake activity in the presence or absence of Mn^{2+} . Thus, it seems that in the presence of Mn^{2+} the donation of electrons by these ions is dominant over the water-donating reaction.

DISCUSSION

Previous communications dealing with photooxidation of Mn^{2+} by chloroplast preparations presented two alternative hypotheses to explain the mechanism of this reaction: (a) direct effect of Mn^{2+} *i.e.* oxidation *via* the electron transport chain, or (b) indirect effect, namely a chemical reaction between Mn^{2+} and some oxidized component produced during illumination, such as H_2O_2 .

In previous studies the effect of Mn^{2+} was tested with either viologens or FMN as electron acceptors. Peroxidase in the presence of Mn^{2+} and H_2O_2 catalyze the oxidation of Mn^{2+} to Mn^{3+} at the expense of the peroxide. An illuminated preparation in the presence of FMN or a viologen under aerobic conditions contains all the components needed for this reaction. Chloroplasts are known to contain a sufficient amount of endogenous peroxidase²⁴, and the product of electron transport mediated by FMN or viologen was shown²⁹ to be H_2O_2 . However, while the reaction of peroxidase is known to be sensitive to either N_3^- or CN^- (ref. 30), the Hill reaction is rather insensitive³¹. This was used as a tool to determine which one of the alternative hypotheses accounted for the Mn^{2+} effect. McKenna and Bishop²⁴ reported that the photooxidation of Mn^{2+} was sensitive to CN^- , though at higher concentrations than required for a soluble catalase system. On the other hand, no such sensitivity could be observed by either Bachofen²³ or by us. Moreover, all our measurements of O_2 uptake in the presence of diquat were carried out in the presence of N_3^- which by itself is a good inhibitor of the peroxidase reaction.

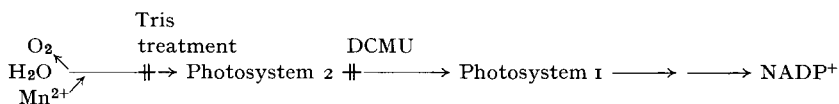
Further evidence pointing to a direct effect of Mn^{2+} came from the experiments with other electron acceptors where H_2O_2 was not formed at all. The best indication was obtained with NADP^+ as the electron acceptor. The results presented in Fig. 3 together with the fact that Mn^{2+} was found to be oxidized by the same system lead to the conclusion that there is a strong competition between Mn^{2+} and water for the donation of electrons which eventually reduce NADP^+ . By analogy, the same picture should have been found for the photoreduction of $\text{Fe}(\text{CN})_6^{3-}$. However, the interaction between the products $\text{Fe}(\text{CN})_6^{4-}$ and Mn^{3+} somewhat obscured the result. Indirect evidence derived from the rate of ATP formation coupled to $\text{Fe}(\text{CN})_6^{3-}$ photoreduction in the presence or absence of Mn^{2+} confirmed the hypothesis that the actual rate of electron flow was not affected by Mn^{2+} . The measured rate was the net rate, reflecting the total rate of $\text{Fe}(\text{CN})_6^{4-}$ production *minus* its reoxidation by the Mn^{3+} formed at the same time. Therefore, the high $P/2e$ ratios observed in the presence of Mn^{2+} are meaningless in regard to the number of coupling sites but rather indicate the existence of a cyclic electron flow induced by the Mn^{2+} .

The DCMU sensitivity of the effect of Mn^{2+} suggested that the site of its action must be close to Photosystem 2. Conclusive evidence for the site of action of Mn^{2+} was drawn from quantum requirement measurements (Table II). The existence of the 'red drop' phenomenon in the Mn^{2+} -induced electron transport must mean that the site of donation of electrons by Mn^{2+} precedes Photosystem 2. Further support

for this conclusion was provided by the interaction between Fe(CN)₆³⁻ photoreduction and Mn²⁺ photooxidation. It was previously shown²⁷ that Fe(CN)₆³⁻ reduction occurs by activation of Photosystem 2 alone. Thus, in order to establish a cyclic flow of electrons as described above, Mn²⁺ must donate electrons prior to Photosystem 2.

Analysis of fluorescence properties of normal and manganese-deficient chloroplast preparations¹⁵ also suggests that the endogenous manganese site of interaction with the electron transport chain is localized on the reducing side of Photosystem 2.

The data presented in this paper could be summarized in the following scheme:



In conclusion, the photooxidation of Mn²⁺ seems to be competitive with the photooxidation of OH⁻ derived from water. This competition explains the increase in the apparent rate of O₂ uptake in the presence of diquat and the inhibition of O₂ evolution in the presence of either NADP⁺ or Fe(CN)₆³⁻. The photooxidation of Mn²⁺ accounts for 70–80 % of the overall activity. It is hard to correlate the activity of added Mn²⁺ with that of the endogenous manganese which is bound to a protein, since no change of the redox state of the latter was reported. It is of interest, that their site of action with respect to Photosystem 2 seem rather similar. Free Mn²⁺ may replace the bound manganese extracted by Tris treatment¹⁵, but they were found to be inactive as electron donors for NADP⁺ photoreduction.

REFERENCES

- 1 A. PIRSON, *Z. Botan.*, **31** (1937) 193.
- 2 A. PIRSON, C. TICHY AND G. WILHELMI, *Planta*, **40** (1952) 199.
- 3 H. C. EYSTER, T. E. BROWN, H. A. TANNER AND S. L. HOOD, *Plant Physiol.*, **33** (1958) 235.
- 4 G. RICHTER, *Planta*, **57** (1961) 202.
- 5 E. KESSLER, *Arch. Biochem. Biophys.*, **59** (1955) 527.
- 6 T. E. BROWN, H. C. EYSTER AND H. A. TANNER, in C. A. LAMB, O. G. BENTLEY AND J. M. BEATLIE, *Trace Elements*, Academic Press, New York, 1958, p. 135.
- 7 G. M. CHENIAE AND I. F. MARTIN, *Brookhaven Symp. Biol.*, **19** (1966) 406.
- 8 D. SPENCER AND J. V. POSSINGHAM, *Australian J. Biol. Sci.*, **13** (1960) 441.
- 9 D. SPENCER AND J. V. POSSINGHAM, *Biochim. Biophys. Acta*, **52** (1961) 379.
- 10 J. V. POSSINGHAM AND D. SPENCER, *Australian J. Biol. Sci.*, **15** (1962) 58.
- 11 B. KOK AND G. M. CHENIAE, in D. R. SANADI, *Current Topics in Bioenergetics*, Vol. 1, Academic Press, New York, 1966, p. 1.
- 12 G. M. CHENIAE AND I. F. MARTIN, *Biochim. Biophys. Acta*, **153** (1968) 819.
- 13 E. KESSLER, W. ARTHUR AND J. E. BRUGGER, *Arch. Biochem. Biophys.*, **71** (1957) 326.
- 14 J. M. ANDERSON AND S. W. THORNE, *Biochim. Biophys. Acta*, **162** (1968) 122.
- 15 P. H. HOMANN, *Biochim. Biophys. Res. Commun.*, **33** (1968) 229.
- 16 P. H. HOMANN, *Biochim. Biophys. Acta*, **162** (1968) 545.
- 17 G. M. CHENIAE AND I. F. MARTIN, *Biochem. Biophys. Res. Commun.*, **28** (1967) 89.
- 18 P. H. HOMANN, *Plant Physiol.*, **42** (1967) 997.
- 19 E. KESSLER, *Planta*, **49** (1957) 435.
- 20 J. M. ANDERSON, N. K. BOARDMAN AND D. J. DAVID, *Biochem. Biophys. Res. Commun.*, **17** (1964) 685.
- 21 R. H. KENTEN AND P. J. G. MANN, *Biochem. J.*, **42** (1949) 255.
- 22 R. H. KENTEN AND P. J. G. MANN, *Biochem. J.*, **61** (1955) 279.
- 23 R. BACHOFEN, *Brookhaven Symp. Biol.*, **19** (1966) 478.
- 24 J. M. MCKENNA AND N. I. BISHOP, *Biochim. Biophys. Acta*, **131** (1967) 339.
- 25 M. AVRON, *Anal. Biochem.*, **2** (1961) 535.
- 26 D. I. ARNON, *Plant Physiol.*, **24** (1949) 1.

- 27 M. AVRON AND G. BEN-HAYYIM, in H. METZNER, *Progress in Photosynthesis Research*, Vol. 3, H. Laupp Jr., Tübingen, 1969, p. 1185.
- 28 M. AVRON AND N. SHAVIT, *Anal. Biochem.*, 6 (1963) 549.
- 29 A. H. MEHLER, *Arch. Biochem. Biophys.*, 34 (1951) 339.
- 30 B. CHANCE, *J. Biol. Chem.*, 194 (1952) 483.
- 31 N. I. BISHOP AND J. D. SPIKES, *Nature*, 176 (1955) 307.

Biochim. Biophys. Acta, 205 (1970) 86-94